The Genotype of the Original Wiskott Phenotype

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Summary

The Wiskott–Aldrich syndrome is an X-linked hereditary disorder associated with combined immunodeficiency, thrombocytopenia, small platelets, eczema, and increased susceptibility to autoimmune disorders and cancers. It is caused by mutations in the gene (WAS) for the Wiskott–Aldrich syndrome protein (WASP). We investigated family members of the patients originally described by Wiskott in 1937 and identified a new frame shift mutation in exon 1 of WAS. This mutation is likely to be the hypothesized genotype that caused the severe form of the Wiskott–Aldrich syndrome in the three brothers described by Wiskott.

In 1937, Alfred Wiskott, a German pediatrician, described three brothers who presented shortly after birth with thrombocytopenia, bloody diarrhea, eczema, and recurrent ear infections; all three died early in life from intestinal bleeding and sepsis. Wiskott commented that “the origin of the hemorrhagic diathesis is a dysfunction in the line of the platelets.” The observation that all three brothers were affected, whereas their sisters showed no symptoms, led Wiskott to propose that the syndrome is due to a “hereditary thrombopathia.” In 1954, Aldrich et al. traced six generations of a family and found that 16 of 40 males, but no females, died of the syndrome first described by Wiskott, thus clearly showing an X-linked mode of inheritance.

The Wiskott–Aldrich syndrome is now known as an X-linked hereditary disorder associated with combined immunodeficiency, thrombocytopenia, small platelets, eczema, and an increased risk of autoimmune disorders and cancers. It has a broad range of phenotypes (Online Mendelian Inheritance in Man no. 301000).

The severe form of the Wiskott–Aldrich syndrome and its milder manifestations — X-linked thrombocytopenia and X-linked neutropenia — are caused by mutations in the gene for the Wiskott–Aldrich syndrome protein (WAS), located at Xp11.22–p11.23 and cloned in 1994. WAS and several related proteins are involved in the reorganization of the actin cytoskeleton by activating the actin-related protein 2/3 complex that mediates actin polymerization in all cells of the hematopoietic system. Mutations in the WAS gene result in truncated or absent WASP in these cells, but there is no strict correlation between the mutant genotype and the expression of WASP or the phenotype of the syndrome. The disorder can be cured through hematopoietic stem-cell transplantation.

We recruited members of the family described by Wiskott in 1937 in order to identify the hypothesized mutation in WAS that caused the severe phenotype of the Wiskott–Aldrich syndrome in the three brothers. Genetic testing for the mutation was carried out in three generations of the kindred.
METHODS

After obtaining written informed consent, we performed mutation analyses in Subjects III-6, III-8, III-9, III-10, IV-2, IV-3, V-1, and V-2 (Fig. 1). Genomic DNA was extracted from peripheral white cells. After amplification, WAS exons 1 through 12 were analyzed with the single-strand conformation polymorphism method and the aberrant fragment of exon 1 was investigated by means of double-strand sequencing. All other WAS exons from Subjects III-10 and V-1 were reanalyzed through direct sequencing (with the use of a kit from Applied Biosystems). The primer sequences used for amplification and the methods used for the screening of mutations have been described previously. A WAS mutation was ruled out in 400 controls (200 men and 200 women) by means of denaturing high-performance liquid chromatography with the Wave system (Transgenomics); the aberrant fragment was observed in DNA from a female carrier, analyzed with the use of buffer B (on a gradient of 53 to 61%) at a running temperature of 64°C. To study the expression of the mutant allele, RNA was isolated from an obligate female carrier and transcribed into complementary DNA (cDNA) with the use of a first-strand cDNA synthesis kit (Amersham Biosciences). The cDNA was sequenced with the use of a pair of primers: one located in the 5’ untranslated region (1cF: 5′TCGCCAGAGAAAGGGCG3′) and one in exon 3 (3cR: 5′CATCTCCAGCGAGGTGTTG3′).

RESULTS

Genetic testing for the mutation revealed a deletion of two nucleotides at positions 73 and 74 in WAS exon 1 (coding sequence, 73–74delAC; the first nucleotide is the A of the ATG translation-initiation codon). This mutation is not listed in WASPbase, an Internet-based database of WAS mutations. The deletion results in a frame shift that starts with amino acid 25; the shifted reading frame is open for another 11 amino acids before it results in a stop codon (protein sequence, Thr25ProfsX12). To further characterize the 73–74delAC mutation in the coding sequence, we first sought it in 400 normal subjects (200 men and 200 women) serving as controls; none carried the mutation. This result makes it improbable that the mutation is a polymorphic variant in the normal popula-
Next, we performed RNA analysis in a female carrier (Subject IV-2). She was heterozygous for the aberrant variant in exon 1. However, after the sequencing of exons 1 through 3 of the generated cDNA, we observed monoallelic expression, which indicates the decay of nonsense-mediated messenger RNA (mRNA) (the destruction of mRNA with a premature stop codon) (Fig. 3). This observation is a sign that the 73–74delAC mutation in the coding sequence is likely to result in the complete absence of WASP in affected men.25

We identified this mutation in three generations of the pedigree: Subjects III-10, IV-2, and V-1. Subject V-1 had presented with symptoms compatible with the severe form of the Wiskott–Aldrich syndrome: bloody diarrhea, severe infections, eczema, and thrombocytopenia with small platelets. He is alive and well after receiving a hematopoietic stem-cell transplant from an HLA-matched unrelated donor (Fig. 2B). Subject III-10, a sister of the three affected brothers described by Wiskott (Subjects III-3, III-4, and III-5), was identified as a carrier of the mutation, but three of her sisters do not carry the mutation. As expected, Subject IV-2, the mother of Subject V-1, also carries the X-linked mutation (Fig. 1).

**DISCUSSION**

Our analysis indicates that Subjects II-2 and II-4 were obligate carriers of the 73–74delAC mutation in the coding sequence after they inherited either a germ-line WAS mutation from one of their parents (Subjects I-1 and I-2) or the mutation from their mother (Subject I-2, the grandmother of Wiskott's patients), who could have been a silent carrier of an ancestral mutation. Since Subjects III-10 and IV-2 are carriers of the mutation, Subject III-2 must be an obligate female carrier.

Our findings indicate the improbability of a spontaneous mutation in Subject V-1 and provide strong evidence that the three affected brothers (Subjects III-3, III-4, and III-5) also had the 73–74delAC mutation in the coding sequence of WAS. Almost 70 years after Wiskott's initial clinical description of the Wiskott–Aldrich syndrome in 1937, we now have a molecular explanation for this rare genetic disorder.
three brothers, we found that a mutation in the X-linked WAS gene caused the severe phenotype. We are indebted to Drs. W. Friedrich and K. Schwarz, Department of Pediatrics, University Hospital Ulm, Ulm, Germany, for providing information about the molecular analysis and stem-cell transplantation performed in Subject V-1, as well as for providing blood samples for additional analysis.

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